Amendments to the specification:

Replace the paragraph found at page 27 (line 19) – page 30 (line 3) of the published PCT publication (WO 01/10911 A2) with the following new paragraph.

Next, the V genes were amplified for cloning into the pCRII-TOPO® vector (TOPO TA-Cloning® Kit, In Vitrogen) for sequence determination. Polymerase chain reaction amplification was done using $V_H$back (5'-CAGGTSMARCTGCAGSAGTCWGG-3') [SEQ ID NO:5] and $V_H$for (5'-TGAGGAGACGGTGACCGTGGTCCCTTGGCCCCAG-3') [SEQ ID NO:6], $V_L$back (5'-GACATTGAGCTCACCCAGTCTCCA-3') [SEQ ID NO:7] and $V_{K2}$for (5'-GGAAGCTTGAAGATGGATACAGTTGGTGCAGC-3') [SEQ ID NO:8] primers with M,R,W and S respectively (A/C), (A/G), (A/T) and (C/T) (all from Eurogentec, Herstal, Belgium) and $V_H$back, $V_H$for and $V_L$back are complementary to the 5'-terminal part of the framework region FR-1 and to the 3'-terminal part of the FR-4 of the V_H- and V_L-genes respectively and V_{K2} for anneals to the C_K sequence. Polymerase chain reactions were performed in a programmable heating block using 30 rounds of temperature cycling (92°C for 1 minute, 55°C for 1 minute and 72°C for 1 minute). The reactions included the cDNA, 1 µg of each primer and 2.5 U of Hotgold polymerase (Eurogentec) in final volume of 50 µl, with the reaction buffers as recommended by the manufacturer (Vitrogen). The polymerase chain reaction product bands were analyzed on a 1.5% agarose gel.